

In the Specification:

Please amend the specification as shown:

Please delete the paragraph on page 18, lines 1-20, and replace it with the following paragraph:

The P4xLuc series of plasmids has been described previously by Deleu et al. in Mol Cell Biol. 18:409-19, incorporated herein by reference (10). P4mut25Luc was cut with PmeI and self-ligated to eliminate a duplication of the NcoI-AflIII fragment within the P4 promoter. P4mut22Luc, P4mut23Luc and P4mut24Luc contain a BglII linker which replaces the E2F, ets and Sp1 sites, respectively. P4mut19Luc and P4mut25Luc contain the linker immediately before the E2F and after the Sp1 sites, respectively. A double stranded oligonucleotide containing four Tcf binding sites (GATC-TCCTTTGATCTTAATCCCTTT GATCTGGATCCCTTTGATCTCCAACCCTTT-GATC) (**SEQ ID NO: 2**) was cloned in both orientations into the BglII site of the P4xLuc plasmids to give plasmids pMM23 & 24 (mut19), pMM25 & 26 (mut22), pMM27 & 28 (mut23), pMM29 & 30 (mut24), and pMM33 & 34 (mut25), where pMM23, 25, 27, 29 and 33 have the C/T-rich strand of the Tcf site on the viral genomic strand and the rest have the Tcf sites reversed. The P4-Tcf promoters obtained were inserted into pH1, a plasmid containing the MVM left hairpin and P4 promoter in the H1 genome (21). To achieve this, the 3298 bp NdeI-EcoRI fragment of pH1 was blunted and self-ligated to give pMM39. The AflIII site at position 3147 in pMM39 was destroyed by self-ligation. The 240 bp AflIII-NcoI fragment of the P4-Tcf-Luc plasmids was cloned into the AflIII-NcoI sites of pMM39 to give pMM41 to 50. Finally, the 2338 bp NheI-SphI fragment of pH1 was cloned into the NheI-SphI sites of pMM41 to 50 to give pMM65 to 74.

Please delete the paragraph on page 19, lines 7-8, and replace it with the following paragraph:

The sequence of parental pH1 virus (21), i.e. H1 genome with MVM P4 promoter and MVM left hairpin as shown in the sequence listing herein (**SEQ ID NO: 1**).

Please delete the paragraph on page 19, lines 11-18, and replace it with the following paragraph:

Cells were harvested and DNA extraction was performed using a Dneasy Tissue kit (Qiagen) according to the manufacturer's instructions. 10 ng of DNA was used per quantitative PCR reaction. TaqMan PCR was performed using a TaqMan Universal PCR Master Mix (Perkin-Elmer), 800 nM primers (Invitrogen) and 500 nM TaqMan probe (MWG) on a PE5700 PCR machine (Perkin-Elmer). The primers and probes lie in the NS1 coding sequence: forward primer, CCACACTCAAAGAGTTGGTACATAA (SEQ ID NO: 3); reverse primer, CACCTGGTTGAGC CATCAT (SEQ ID NO: 4); probe, AACTGTCTGGCTGCATCATCATCCA (SEQ ID NO: 5).